# Mesenchymal Stem Cells on Horizon: A New Arsenal of Therapeutic Agents

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#### Abstract

Over 10 years, mesenchymal stem cells (MSCs) have been considered as valuable and suitable cells for cell-based therapy applications, particularly in clinical trials. In any case, they are as yet not utilized routinely in clinics. At first, it was believed that MSCs play their roles, especially in regenerative medicine due to their differentiation and cell replacement properties. Interestingly, it is well-known that MSCs mainly exert their therapeutic effects through their vast bioactive factors. These findings turned scientists' consideration toward cell-free therapy concepts. From this point of view, MSCs can be considered as an arsenal of natural bioreactors in variety of therapeutic agents. MSCs inherently express various important therapeutic agents such as growth factors and cytokines that can be manufactured, handled and stored as a prepared-to-go biologic product. In this review, we provide a vision, highlight as well as discuss in order to introduce competitive natural robust bioreactor MSCs on the horizon.

Keywords MSC · Secretome · Cell-free therapy · Condition medium · Therapeutic agents

## Introduction

Mesenchymal Stem Cells (MSCs) are multipotent stem cells that have a well-defined capacity for self-renewal [1]. MSCs express CD73, CD105, and CD90, while they have no expression of CD34, CD45, CD14 or CD11b, CD19a, and HLA-DR surface markers. Moreover, they have the capacity to differentiate into adipocytes, osteoblasts, and chondroblasts *in vitro* [2, 3]. Over the past decade, MSCs have gained much more attention in regenerative medicine area because of homing ability, immune regulatory properties [4], lower ethical concerns, and tumorigenicity [5], as well as, transdifferentiation capacity [6], tissueorgan repairing, and promoting the survival of damaged tissues

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[7]. In addition, MSCs-based therapies have been performed in a number of trials worldwide [8] (https://clinicaltrials.gov).

In spite of many advantages of MSCs for cell therapy purposes, there are several challenges for clinical use. Some of them are; uncontrolled cell quality, invasive cell isolation process, loss of potency, limited lifespan, the gradual loss of their initial properties during expansion and in vitro proliferation. It is noteworthy that, stressful conditions such as oxidative stress, serum deprivation, inflammation, chemotherapy and radiotherapy in the recipients' tissues/organs, decrease the cell's survival dramatically, resulting in the death of around 99% of MSCs during a few days after transplantation [9–18]. Inadequate quantity of transplanted cells is another remarkable limitation of MSCs application. Noteworthy, more than one hundred million of MSCs require for cell therapy and it takes about ten weeks to prepare sufficient cells before transplantation. Furthermore, clinical features and age of patients are other concerns which influence on the optimal culture conditions of MSCs [19, 20]. In other words, aging and senescence phenotype in MSCs are other important limitations for clinical use. It has been shown that MSCs from aged-patients exhibit characteristics of aging and senescence such as epigenetic modifications, DNA mutations, mitochondrial dysfunction, and telomere length [21]. It has also been revealed that growth kinetic of adipose-derived stromal/stem cells (ASCs) is positively correlated with the donor's age. The proliferation of MSCs decreased in elderly



people whereas apoptosis increased. Moreover, it seems that the differentiation potentials of MSCs change with increased age [16, 22, 23]. These observations indicate a necessary efficient solution to rejuvenate MSCs in vitro when clinical applications of them are considered.

Moreover, some evidence indicating that MSCs have a cancerous origin in the body tissues [24, 25]. In other words, there are common characteristics between MSCs and cancer stem cells that may result in tumorigenesis, which make them unsuitable for direct utilize in the clinic.

Furthermore, the effect of metabolic disorders on MSC's fate could be other limitation of them in clinical application. In other words, metabolic disorders may cause an effect on quality of MSCs. It has been shown that MSCs harvested from equine with metabolic disorders had lower proliferation rate, mitochondrial dysfunction, and higher autophagy cell death in comparison with healthy equine. Overall, the results of this study indicated that autologous MSCs transplantation could be challengeable in patients who are suffering from metabolic disorder [26].

On the other hand, a body of studies indicates that the therapeutic properties of MSCs are owing to their paracrine effects of growth and nutritional factors. Other studies have also shown that the stem cell-derived secreted agents are able to exert therapeutic effects without presence of any other cells [27–29]. MSCs' secretory trophic factors, hormones, and cytokines are known as secretome that can be produced in

environment which stem cells are grown; which named that, conditioned medium (CM) [4] (Fig. 1). Note that, exosomes are part of the MSC secretome.

Therefore, in recent years, MSCs-derived secretome has been introduced as a promising candidate for novel cell-free therapy. For example, Camussi et al. reported that MSC's secretome prohibited kidney injury [30, 31]. In addition, it has been shown that the MSC exosomes of mice exert therapeutic effects to improve pulmonary hypertension in lung tissue [32, 33]. Other studies also indicate the MSC's secretome therapy promoted re-epithelialization of cutaneous wounds by inducing epithelial cell proliferation [34] and angiogenesis [35]. Several studies have been shown the presence of cytokines, hormones and growth factors in MSCs-derived CM which resulted in repairing of damaged tissues [36–45].

Here, we are going to introduce MSCs as an arsenal of therapeutic, beneficial and high-performance agents. In other words, we discuss and highlight the presence of various growth factors/cytokines and tissue regenerative factors, that making the MSCs as a natural, valuable, promising and versatile bioreactor in order to produce pharmaceuticals agents.

### Secretome as a Novel Approach for Cell-free Therapy

chemokines, cytokines, growth factors and extracellular

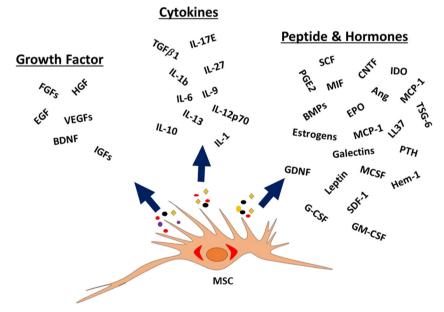
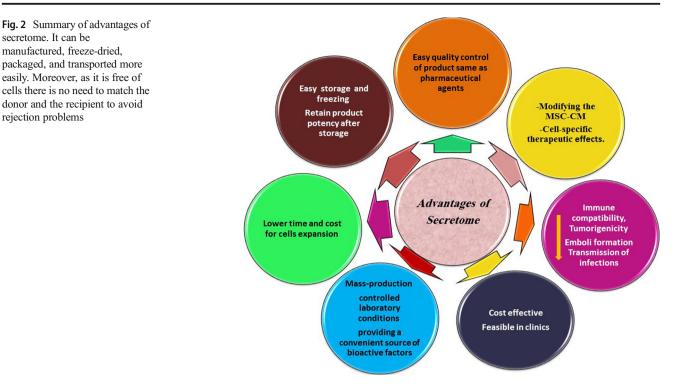


Fig. 1 MSCs secretory trophic factors, hormones, and cytokines are known as secretome. Growth factors: BDNF; Brain-Derived Neurotrophic Factor, EGF; Epidermal Growth Factor, FGFs; Fibroblast Growth Factor, HGF; Hepatocyte Growth Factor, IGFs; Insulin-Like Growth Factor, VEGF; Vascular Endothelial Derived Growth Factor. Cytokines: TGF-B1; Transforming Growth Factor Beta, IL-6; Interleukin 6, IL-10; Interleukin 10, IL-27; Interleukin 27, IL-17E; Interleukin 17E, IL-13; Interleukin 13, IL-1Ra; Interleukin 1 receptor antagonist, IL-8;Interleukin 8, IL-9; Interleukin 9, IL-1b; Interleukin-1b.

Peptide & hormones: Ang; Angiopoietin, BMPs; Bone Morphogenetic Proteins, CNTF; Ciliary Neurotrophic Factor, EPO; Erythropoietin, GDNF; Glial cell line-Derived Neurotrophic Factor, G-CSF; Granulocyte Colony Stimulating Factor, GM-CSF; Granulocyte Macrophage CSF, MCSF; Macrophage CSF, HO-1; Hemoxygenase-1, IDO; Indoleamine 2,3-Dioxygenase, MCP-1; Monocyte chemotactic Protein, MIF; Macrophage Migration Inhibitory Factor, PGE2; Prostaglandin E2, SCF; Stem Cell Factor, SDF-1; Stromal Cell-Derived Factor 1, TSG-6; (TNF)-Stimulated Gene-6, LL37

MSCs have the ability in order to produce a wide range of



matrix (ECM) molecules. The MSC's niche including a variety of microenvironmental signals which are generated during healing, development or diseases, in turn, it regulates tissue regeneration through proliferation and differentiation [46]. The secretome is defined as a series of molecular factors which is secreted into extracellular space. These factors consist of hormones, soluble proteins, cytokines and growth factors [47]. The scientific evidence indicates that similar to the cellular counterpart, MSC's secretome can be used in order to exert favorable effects in tissue regeneration [48]. In other words, cytokines and growth factors produced in MSCs can be used for cell-free regenerative medicine. Interestingly, each cytokine and growth factor can be considered as a novel potential therapeutic agent [48]. Therefore, it may have a significant impact in the near future. Table 1 indicates the studies dealing with the usage of CM for treatment of a variety of diseases. However, depend on the tissues that MSC's CM/secretome have been isolated the contents of them are variable.

### Advantages of Secretome as a Therapeutic Agent

As previously mentioned, the secretome is cell-free. Therefore, in allogeneic usage, it will reduce the risk of adverse reactions. On the other hand, secretome's therapeutic doses can be achieved with one million of MSCs. Secretome can be stored at -80 °C without any significant loss of quality and function which is ready to use after thawing immediately [50, 61]. In terms of stability, some growth factors/cytokines such as IL-6 are stable; IFN- $\gamma$  and MIP-2 are somewhat stable at 4 °C and TNF- $\alpha$ , IL-10 and IL-17A are not stable in

secretomes. Therefore, secretome containing IL- 10 and IL-17A can be stored at -80 °C; however, it is recommended to measure after the first thaw [62]. However, regarding with stability of secretome further studies are required.

The secretome-based therapy can be performed at the regular intervals for a long time, for example at weekly intervals, providing therapeutic courses instead of a single therapeutic at only one- time point. Figure 2 shows some advantages of secretome in regenerative medicine [50, 61, 62].

### Secretome-based Therapy in Regenerative Medicine

A number of studies have been shown that secretome contains of immune-modulatory, anti-inflammatory, anti-apoptotic, anti-oxidant, anti-fibrotic, anti-bacterial and neuroprotective properties. Therefore, it can be employed in a variety of diseases. Moreover, it represents a ready-to-use therapeutic agent. Table 2 shows some current therapeutic applications in regenerative medicine. It is noteworthy that the majority of studies dealing with employing of secretome are in preclinical studies. Overall, these studies indicate that secretome, like. MSCs, is applicable for curing many of diseases.

### MSCs as a Natural Arsenal of Therapeutic Agents

The various secretory agents produced by MSCs in the microenvironment could have a therapeutic potential. These secretory agents include growth factors, proinflammatory and anti-inflammatory cytokines, as well as other peptides and hormones. As mentioned before, it's

Table 1 Summa	try of studies in whic	ch MSCs-derived conditic	Summary of studies in which MSCs-derived conditioned medium (MSCs-CM) are employed to treat in a variety of diseases	ed to treat in a variety of diseases	
Diseases	Donor cells- derived CM	Clinical trials or preclinical	Rout of injection	Outcome	References
Spinal cord injury	BMMSCs	Preclinical/ Rat	Directly injection	<ul> <li>Increased angiogenesis</li> <li>Protected neurons from apoptosis</li> <li>Improved functional recovery after SCI <i>in vivo</i></li> </ul>	[49]
Corneal epithelial wound healing	hUCESCs	Preclinical / Rat	Topically (1 drop)	- recurect tysue cavity <i>in 1100</i> - Anti-inflammatory effects -Bactericidal effects	[36]
wound incaning Uveitis	hUCESCs	Preclinical / Rat	Topically (1 drop)	-Limaneus conteat would heating -Reduced leucoytes Leednord ocnitar inflammation	[50]
Acute and chronic hind limb ischemia	ADSCs	Preclinical / Mouse	Intramuscular	-Reduced apoptosis - Reduced apoptosis -Inhancement of endothelial cell growth -Improvement of blood perfusion in the hind	[37]
Cerebral injury	BMMSCs	Preclinical / Sprague- Dawley Rat	Intravenous	-Attenuated brain damaged volume - Attenuated incidence during TBI (traumatic brain injury) -Decreased TBI-induced neuronal apoptosis	[51]
Lung injury	BMMSCs	Preclinical / Mouse	Intratracheally	<ul> <li>Increased neurogenesis (under normoxic or hypoxic conditions)</li> <li>Decreased LPS-induced lung injury with soluble factors IGF-I</li> <li>Decreased lung inflammation</li> </ul>	[52]
Bone defects	BMMSCs	Clinical	Surgical/ Maxillary sinus floor elevation (SFE) (lateral window ammosch)	<ul> <li>Decreased tung vascual permeabury</li> <li>Increase bone volume in the center of the augmented area -Clinically confirmation of [53] bone formation in all cases</li> </ul>	[53]
Colitis	Amniotic fluid	Preclinical / Mouse	Intraperitoneal	- Decreased the inflammation - Decreased levels of MMP2 protein	[54]
Alopecia	ADSCs	Preclinical/ Nude Mice	Subcutaneous	-increased revels of 10001 -Hair growth stimulation effects	[39]
Muscular	ADSCs	Preclinical/ Rat	Local / systemic injection	-increased growin factor secretion in hypoxic CM -Decreased attrophy of the rotator cuff muscles Decremental minoral attractions	[55]
Myocardial infarction	ADSCs	Preclinical/ Pig	Intravenous	-revented muscie degeneration. -Improved Systolic and diastolic cardiac performance -Reduced movtocardial oxidative stress -Reduced anortocis	[56]
Acute liver injury/ failure	Amniotic fluid	Preclinical/ Nude Mice Intrahepatically	Intrahepatically	-Induced liver recovery with presence of anti-inflammatory factors	[38]
Skin wound	ADSCs	Human clinical study /Original	Topically	<ul> <li>Reduced erythema and pigmentation</li> <li>Fasten recovery of the skin barrier function</li> <li>Enhancing wound healing</li> </ul>	[57]
	PDLSCs	Preclinical/ Nude Mice Intravenous	Intravenous		[58]

Table 1 (continued)	nued)				
Diseases	Donor cells- derived CM	Clinical trials or preclinical	Rout of injection	Outcome Refe	References
Multiple sclerosis				<ul> <li>-Augmented neurons spine density and remyelination</li> <li>-Induced anti-inflammatory and immunosuppressive effects</li> <li>- Attenuated apoptosis -Immunosuppressive effects</li> <li>-Protected against EAE (experimental autoimmune encenhalonwolitis)</li> </ul>	
Parkinson's disease	WJMSCs	Preclinical/ Rat	Directly into the medial forebrain bundle (MFB) - Impact on brain structure and animal behavior - Restored the Neuronal Structure in PD	[59]	[6
Atrophied muscles	UCPVCs	Preclinical/ Rat	Intramuscular	-Suppressed atrophy-related ubiquitin E3-ligases, MuRF-1, and MAFbx. - Regenerated muscle mass and proteins in atronhied muscles.	[0]
Cancer	hUCESCs	Preclinical/ Mouse	Intratumorally	-Modified cell cycle: increased G0-G1 phase, Modified cell cycle: increased G0-G1 phase, decreased G2-M, decreased expression of cyclin A, cyclin B, and cyclin D1 proteins -Induced apoptosis in the MDA-MB-231 cell line -Inhibited invasion, 3D growth, and tumor volume - Reduced proliferation in breast tumors	[[
BMMSCs: Bor hUCESCs: Hur	ne marrow-derived n nan uterine cervical (	BMMSCs: Bone marrow-derived mesenchymal stem cells; ADSC: hUCESCs: Human uterine cervical stem cells; UCPVCs: Umbilical	ADSCs: Adipose-derived stem cells; DPSC: Dental pulp stem cells; W. bilical cord perivascular cells; PDLSCs: Periodontal ligament stem cells	BMMSCs: Bone marrow-derived mesenchymal stem cells; ADSCs: Adipose-derived stem cells; DPSC: Dental pulp stem cells; WJMSCs: Umbilical cord Wharton's Jelly mesenchymal stem cells; hUCESCs: Human uterine cervical stem cells; UCPVCs: Umbilical cord perivascular cells; PDLSCs: Periodontal ligament stem cells	tem cells;

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Table 2         Applications of secretome therapy	stome therapy			
Disease	Tissue/Model	Outcome	Source of CM/Target	Reference
Circulatory system injury	Heart/Infarct, IR	-Cardio protective activity -Effective in animal models of mvocardial infarction	-Human /Pig -Human /Mouse -Rat/Rat	[56, 63, 64]
Nervous system injury	Brain/Stroke, Ischemia	-Effective in stroke -Effective in peri-natal hypoxic-ischemic brain injury -Effective in hind-limb ischemia -Exhibited potent neuro protective activities in neurons - Models of reanmatic bein intury.	-Rat /Rat -Human /Ovine -Human /Mouse	[65–67]
Digestive system injury	Liver/fibrosis	-Amount of the second second much of the second sec	-Human /Mouse -Human /Rat -Rat /Rat	[68–70]
Respiratory system injury	-Lung / hypoxia, E.coli endotoxin, silicosis, fluid filled	- Effective in improving pulmonary hypertension - Effective in improving pulmonary hypertension - Effective in improving silicosis - Cleared shoolar fluid from human humes av vivo	-Mouse /Mouse -Human /Mouse - Human /Human	[33, 71–73]
Skin and subcutaneous tissue injury	Skin /wound	<ul> <li>-Created arrow number number number of several arrows arrow arrow number of cutaneous wounds by inducing epithelial cell proliferation</li> <li>-Activated collagen and elastin secretion by fibroblasts</li> <li>-Prevented modefluxohlast formation thereby reducing secrition</li> </ul>	-Human /Rat -Human /Mouse	[34, 74, 75]
Musculoskeletal system and connective tissue initry	-Skeletal Muscle/cardiotoxin	Promoted muscle regeneration	-Human /Mouse	[76, 77]
Acute kidney injury (AKI)		-Attenuated renal injury -Improved kidney recovery competence -Attenuated necrosis, apoptosis, and inflammation -Increased cellular proliferation	-Human /Pig - Human /Mouse - Human /Rat	[78–80]

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Abbreviation	Full name	Functions	Reference
Growth factor			
BDNF	Brain Derived Neurotrophic Factor	Promotes survival and differentiation of neurons, reduces infarct size	[49]
EGF	Epidermal Growth Factor	Induces cell proliferation and differentiation	[38, 39, 41, 42]
FGF	Fibroblast Growth Factor 2/Basic Fibroblast Growth Factor (FGF-2/BFGF)	Induces angiogenesis, inhibit apoptosis	[16, 37, 41, 42, 86]
HGF	Hepatocyte Growth Factor	Promotes progenitor cell mobilization, induces angiogenesis and cell proliferation, inhibits immune cell proliferation	[37, 38, 45, 87, 88]
IGF	Insulin-Like Growth Factor I (IGF-I) Insulin-Like Growth Factor II (IGF-II)	Induces cell proliferation, inhibits apoptosis	[39, 88, 89]
KGF/FGF-7	Keratinocyte Growth Factor/Fibroblast Growth Factor 7	Induces cell proliferation	[38, 42]
VEGF	Vascular Endothelial Derived Growth Factor	Induces angiogenesis, promote progenitor cell mobilization, inhibits apoptosis	[16, 37, 39, 41, 42, 87, 90]
Abbreviation	Full Name	Functions	
Anti-inflammatory cytokines	tines		
$TGF - \beta 1$	Transforming Growth Factor $\beta$	Induces stem cell differentiation, reduces inflammation/ immune activation	[38, 42, 88, 91]
IL-6	Interleukin 6	Stimulates stem/progenitor cell proliferation, induces angiogenesis	[16, 40, 41, 45, 49, 89, 90]
IL-10, IL-27, IL-17E, Int IL-13, IL-12p70, and (IL-1ra) Pro-inflammatory cytokines	Interleukin 10, Interleukin 27, Interleukin 17, Interleukin 13, Interleukin 1 receptor antagonist ines	Anti-inflammatory	[38]
IL-8	Interleukin 8	-Involves in mitogenesis -Inhibits angiogenesis, inflammation, chemotaxis, neutrophil degranulation, leukocyte activation, and calcium homeostasis.	[40, 41, 87]
IL-9	Interleukin 9	Stimulates cell proliferation and prevents apoptosis.	[41, 92]
IL-1b	Interleukin-1b	suppresses inflammation	[38]
Peptide & Hormones			
Ang	Angiopoietin	Induces angiogenesis and promotes cell survival	[87]
BMP	Bone Morphogenetic Protein	Regulates tissue homeostasis, promotes neurogenesis, induces stromal cell proliferation and migration, promotes anziogenesis	[88]
CNTF	Ciliary Neurotrophic Factor	Promotes survival of neurons	[93]
EPO	Erythropoietin	Induces angiogenesis, inhibits apoptosis	[94]
Gal	Galectins	Suppresses inflammation, induces stem cell mobilization, inhibits immune cell proliferation	[95, 96]
		-	

Table 3Some trophic factors and cytokines produced by MSCs and suggested functions for tissue regeneration/repair

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Table 3 (continued)			
Abbreviation	Full name	Functions	Reference
GDNF	Glial cell line-Derived Neurotraphic Factor	Promotes survival of neurons, induces axonal growth, reduces infarct size	[94]
G-CSF	Granulocyte-Colony Stimulating Factor	Induces stem/progenitor cell proliferation, promotes neuronal differentiation	[39, 42]
GM-CSF	Granulocyte Macrophage- Colony Stimulating Factor	Stimulates stem cells to produce granulocytes and monocytes, plays a role in embryonic development, as a vaccine adjuvant in HIV-infected nations	[39, 41, 42]
M-CSF	Macrophage- Colony Stimulating Factor	Induces the proliferation, differentiation, and survival of monocytes, macrophages, and bone marrow progenitor cells, Increases tumor cell evitoxicity	[39]
HO-1	Heme Oxygenase-1	Promotes induction of regulatory T cells	[67]
IDO	Indoleamine 2,3-Dioxygenase	Induces regulatory T cells, inhibits T cell activation	[97, 98]
MCP-1	Monocyte Chemotactic Protein 1	Induces angiogenesis, induces MSC migration, inhibits apoptosis	[16]
MIF	Macrophage Migration Inhibitory Factor	Inhibits macrophage migration	[38, 40, 41, 90]
PGE2	Prostaglandin E2	Suppresses inflammation, inhibits immune cell proliferation	[66]
SCF	Stem Cell Factor	Induces stem/progenitor cell proliferation, promotes neuronal differentiation	[97, 100]
SDF-1	Stromal Cell-Derived Factor 1	Regulates progenitor cell mobilization	[16, 37, 38, 87]
TSG-6	(TNF)-Stimulated Gene-6	Suppresses immune activation	[101, 102]
LL37	Human cathelicidin antimicrobial peptide	Anti-bacterial effect of MSC, direct killing of microorganisms, chemotaxis and chemokine induction	[103, 104]
		-Regulates inflammatory responses - Angiogenic, anti-apoptotic, and wound healing effects	
Leptin	1	Promotes adipogenesis and reduces osteogenesis	[105]
Estrogen	1	Involves in memory impairment, increases fat store, maintenance of vessel and skin, reduces bone resorption, increases bone formation, supports hormone-sensitive breast cancers, promotes lung function by supporting alveoli	[106]

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identified most of them with a general name. "secretome". It can be injected by several ways: 1) direct injection into damaged tissues 2) intravenously, 3) transdermal and 4) intramuscular. The different proteomic studies have shown that various agents are present in secretome [31, 63, 81–85]. In fact, if MSCs consider as "building blocks", their cytokines, growth factors and hormones can be assumed as "workers". Growth factors lead to the activation, stimulation, and mobilization of stem cells from their origin. If more specific growth factors are produced, it would be possible in order to regenerate the damaged tissue specifically. Table 3 shows some cytokines, growth factors and hormones which are secreted by MSCs and their potential functions for tissue regeneration/repair.

# Cytokines, Growth and Soluble Factors Act as Immunomudulatory

MSCs interact with various lymphocytes. During the innate and acquired immune systems, MSCs are able to inhibit the activation of pro-inflammatory monocytes and macrophages using secreting soluble factors such as transforming growth factor- $\beta$  (TGF- $\beta$ ), hepatocyte growth factor (HGF), nitric oxide (NO), heme oxygenase (HO), interleukin (IL)-6, prostaglandin E2 (PGE2), indoleamine 2, and 3-dioxygenase (IDO) [107, 108]. Based on preclinical studies, it has been shown that MSCs have suppressive effects on both adaptive and innate immunity systems [109, 110]. MSCs are able to inhibit the activation of pro-inflammatory monocytes and macrophages. Moreover, in the presence of MSCs and their soluble factors, monocytes and macrophages may acquire antiimmunosuppressive functions. Moreover, MSCs inhibit the differentiation of monocytes into fully matured dendritic cells (DCs). The tolerogenic DCs produce a high level of IL-10 and decrease the ability of stimulate allogeneic T-cell proliferation in a mixed lymphocyte reaction [111–118]. MSCs prevent the proliferation and cytotoxicity of natural killer cells (NKs) mainly through PGE2 and IDO productions. MSCs are also able to suppress T-cell proliferation through the secretion of various soluble factors and inhibit T-cell activation. The immune modulatory factors are summarized in fig. 3. In addition, extensive clinical trials dealing with the immunomudolatory contribution of MSC-derived growth factors/ cytokines have also been reported (http://clinicaltrials.gov). MSC-derived growth factors/ cytokines have been promising for treatment of graft versus host disease(GVHD) [119, 120], inflammatory and autoimmune diseases such as multiple sclerosis or Crohn's disease, diabetes mellitus type I and systemic lupus erythematosus (SLE) [110]. Pretreatment of MSCs with IFN- $\gamma$  resulted in preventing of GVHD [121, 122].

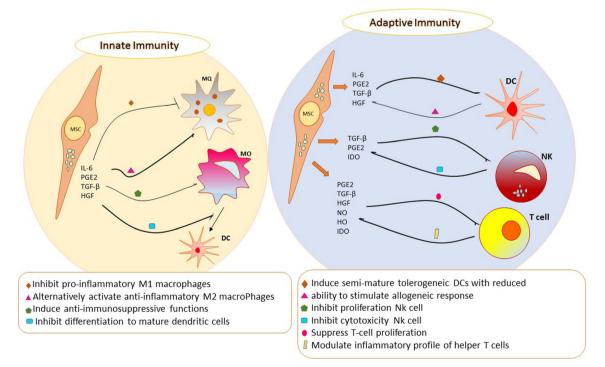


Fig. 3 Immunomodulatory properties of MSC-secretome. MSCs interact with various lymphocytes. During the innate and acquired immune systems, MSCs are able to inhibit the activation of pro-inflammatory monocytes and macrophages through the secretion of soluble factors. Additionally, MSCs inhibit the differentiation of monocytes into fully matured dendritic cells (DCs). MSCs prevent the proliferation and cytotoxicity of natural killer cells (NKs) and suppress the proliferation of the T cell. They also prevent the activation of the T cell through the cell to cell contact. MQ; Macrophage, MO; Monocytes, DC; Dendritic cells, NK; Natural killer

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Tissue/ disease	Growth factor/cytokine	Reference			
Neutropenia - G-CSF [128–130] - GM-CSF Wound healing - EGF [131, 132] - TGF-B + HGF - VEGF - PDGF - PDGF - FOF-L&II - TGF-C - KGF Cardiovascular & - VEGF [133, 134] Infarction - TNF - IL-6 - LR-8 - MCP-I - HGF - GM-CSF - SDF-1 and receptor CXCR4 - VEGF - MCP-1, 3 - Ang-1 - NO - SDF-1a - CSF - GM-CSF - GM-CSF - GM-CSF - CSF - GM-CSF - CSF - CSF	Bone	- IGF-II - TGF-B - FGF - PDGF - PTHRP - BMP - GDF	[123–127]		roduct Name	nterferon $\alpha$ -2a, recombinant
Wound healing $= EGF$ [131, 132] = TGF-B = HGF = VEGF = PDGF = FGF-I&III $= TGF-\alpha$ = KGF $= TGF-\alpha$ = KGF = TMP-2 Infarction $= TSP-1$ = TFF = L-6 = L-8 = MCP-1 = HGF = CGF-2 = LF = SCF = G-CSF = G-CSF = G-CSF = GGF-7 = EGF = TGF-1 = TGF-2 = LF = SCF = G-CSF = GGF-7 = EGF = TGF-7 = EGF = TGF-7 = EGF = TGF-7 = EGF = TGF-7 = EGF = TGF-8 = 1GF = TGF-8 = 1GF = TGF-7 = EGF = TGF-8 = 1GF = CGF = CGF	Neutropenia	- G-CSF - GM-CSF	[128–130]			
$ \begin{array}{c} Cardiovascular & -VEGF [133, 134] \\ \mbox{Myocardial -TIMP-2 \\ Infarction -TSP-1 \\ -TSP-1 \\ -TSP-1 \\ -TNF \\ -IL-6 \\ -IL-8 \\ -MCP-1 \\ -HGF \\ -FGF-2 \\ -LIF \\ -SCF \\ -GCSF \\ -GCSF \\ -GM-CSF \\ -EPO \\ -IGF-1 \\ -SDF-1 (SDF-1/CXCL12) \\ -Ang-1 \\ -TNF-\alpha \\ \\ Liver +HEGF \\ -HGF \\ $	Wound healing	- EGF - TGF-B - HGF - VEGF - PDGF - FGF-1&II - TGF-α	[131, 132]		Brand	Roferon®-A
FGF-7 - EGF - HGF - TGF-B - IGF - SDF-1and receptor CXCR4 - VEGF Cardiovascular - TGF-B - IGF-B - IGF - SDF-1and receptor CXCR4 - VEGF - MCP-1,3 - Ang-1 - NO - SDF-1a - CSF - CSF - CSF - CMCSE	Myocardial	- VEGF - TIMP-2 - TSP-1 - TNF - IL-6 - IL-8 - MCP-I - HGF - FGF-2 - LIF - SCF - G-CSF - GM-CSF - EPO - IGF-1	[133, 134]		properties	
FGF-7 - EGF - HGF - TGF-B - IGF - SDF-1and receptor CXCR4 - VEGF Cardiovascular - TGF-B - MCP-1,3 - Ang-1 - NO - SDF-1a - CSF -		- Ang-1 - TNF-α 			nical use	airy Cell Leukemia Malignant Melanoma ollicular Lymphoma
- SDF-1a - CSF	Liver	- FGF-7 - EGF - HGF - TGF-B - IGF - SDF-1and receptor CXCR4	[38, 135]	roved by FDA	Cli	eered <i>E. coli</i> strain / -H <sup>E</sup> iman leukocytes - M - F <sub>0</sub>
- IL-6	Cardiovascular	- TGF-B - VEGF - MCP-1,3 - Ang-1 - NO - SDF-1a - CSF - GMCSF	[136–142]	ytokines/growth factors appr		Genetically engine IFN-α from hu
Table 5 Cytokin IFN-α				Table 5 C <sub>3</sub>	cine/gro	~

Table 4 Tissue- specific MSC-derived growth factors/cytokines impli-

Recombinant human platelet-derived growth factor (rhPDGF-BB)

granulocyte-macrophage colony stimulating factor (rhGM-CSF)

Recombinant human

Leukine (sargramostim) (becaplermin) Gel

- Acute Myelogenous Leukemia For injection, intravenous infusion
 - Mobilization and Engraftment (Rx Only)

- Autologous and allogeneic bone of PBPC (peripheral blood

progenitor cells)

marrow transplantation

Human recombinant interleukin-2

**PROLEUKIN®** (aldesleukin) REGRANEX

For injection, intravenous infusion

Genetically engineered E. coli strain -Metastatic renal cell cancer

IL-2

- Condylomata Acuminata

For external use only (Rx Only)

Lower extremity diabetic

Yeast, Saccharomyces cerevisiae Yeast, Saccharomyces cerevisiae

rh-GM-CSF PDGF-BB

neuropathic ulcers

- Metastatic melanoma - Chronic Hepatitis C

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### **Tissue- Specific MSC-derived Factors**

The various factors may be presented into secretome as a cocktail and act in concert in order to promote regeneration. Therefore, it is important to analyze a complete set of growth factors and cytokine levels for every kind of stem cell-derived secretome/ conditioned medium. While the content of the various cytokines in a certain secretome/conditioned medium is known, the potential outcome can be increased and translated to targeted therapy in regenerative medicine [45]. For example, bone regeneration could be occurred in the presence of insulin-like growth factor (IGF-I and IGF-II), transforming growth factor beta (TGF- $\beta$ ), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), parathyroid hormone-related peptide (PTHRP), bone morphogenic protein(BMP) into secretome/conditioned medium. Furthermore, the presence of epithelial growth factor (EGF), TGF-  $\beta$ , hepatocyte growth factor (HGF), vascular endothelial derived growth factor (VEGF), PDGF, FGF-I&II, TGF- $\alpha$ , and keratinocyte growth factor (KGF) play an important role in wound healing. Some of the specific growth factors and cytokines, which are implicated in the regeneration and repair of any tissue/disease, are listed in Table 4.

# Growth Factors, Cytokines, and Hormones Available in the Market

The emergence of new classes of therapeutic agents which manufactured by biotechnology is one of the more exciting

Table 6Mesenchymal stem cell recombinant growth factors

Product	Quality grade	Description	Source
Human FGF-1	Research grade	Recombinant human fibroblast growth factor 1	E. coli
Human FGF-2	Research grade	Recombinant human fibroblast growth factor 2	E. coli
Human BDNF	Research grade	Recombinant human brain-derived neurotrophic factor	E. coli
Human BMP-2	Research grade & Premium grade	Recombinant human bone morphogenetic protein 2	E. coli
Human BMP-4	Research grade & Premium grade	Recombinant human bone morphogenetic protein 4	Pichia pastoris
Human BMP-6	Research grade	Recombinant human bone morphogenetic protein 6	HEK293 cells
Human BMP-7	Research grade	Recombinant human bone morphogenetic protein 7	CHO cells
Human EG-VEGF	Research grade	Recombinant human endocrine gland-derived vascular endothelial growth factor	E. coli
Human EGF	Research grade & Premium grade	Recombinant human epidermal growth factor	E. coli
Human FGF-2 IS	research grade	Recombinant human fibroblast growth factor 2 IS (improved sequence)	
Human G-CSF	research grade premium grade	Recombinant human granulocyte colony- stimulating Factor	E. coli
Human Galectin-1	research grade	Recombinant human galectin 1	E. coli
Human GM-CSF	research grade premium grade	Recombinant human granulocyte macrophage colony-stimulating factor	E. coli
Human HGF	Research grade	Recombinant human hepatocyte growth factor	E. coli
Human IFN-α2a Human IFN-α2b	Research grade	Recombinant human interferon $\alpha 2a$ Recombinant human interferon $\alpha 2b$	E. coli
Human IFN-β1a	Research grade	Recombinant human interferon $\beta$ 1a	CHO cells
Human IFN-β1b		Recombinant human interferon $\beta$ 1b	E. coli
Human IFN-γ1b	Research grade	IFN- $\gamma$ Recombinant human interferon $\gamma$ 1b	E. coli
Human IGF-1 Human IGF-2	Research grade	Recombinant human insulin-like growth factor 1 Recombinant human insulin-like growth factor 2	E. coli
Human M-CSF	Research grade	Recombinant human macrophage-colony stimulating Factor	E. coli
Human MCP-1	Research grade	Recombinant human monocyte chemotactic protein 1	E. coli
Human MIF	Research grade	Recombinant human macrophage migration inhibitory Factor	E. coli
Human SCF	Research grade premium grade	Recombinant human stem cell factor	E. coli
Human SDF-1a	Research grade	Recombinant human stromal cell-derived factor $1\alpha$	
Human TGF-β1 Human TGF-β2 Human TGF-β3	Premium grade Research grade	Recombinant human transforming growth factor $\beta 1$	HEK293 cells Insect cells HEK293 cells
Human TNF- $\alpha$	Premium grade Research grade	Recombinant human tumor necrosis factor $\alpha$	E. coli Yeast
Human VEGF	Research grade	Recombinant human vascular endothelial growth factor	Insect cells

frontiers in medicine. The possibility of clinical usage of recombinant growth factors and cytokines expressed by MSCs has been proved; however, the application of that is limited. Reproducibility of the growth factors, cytokines, HLA incompatibility and the infectious agentstransmission possibility might be the reasons behind them. The emergence of MSCderived growth factors and cytokines will address many of these problems and pave the way for its evaluation in a variety of diseases. A number of cytokines have already been licensed by the Food and Drug Administration (FDA) in clinical application (Table 5). The FDA must evaluate an ever-increasing number of new growth factors and cytokines. In order to develop regulatory policy for using of these products from laboratory bench to the bedside, several factors including those combined sound scientific principles and good clinical medicine should be considered, however, the final goal should be beneficial for patients. In order to achieve a successful cellbased clinical trial, high quality of raw materials is essential. Interestingly, the MSCs inherently express many growth factors and cytokines with the highest degree of good manufacturing practice (GMP). Therefore, harvesting these therapeutic agents from MSCs would be an important part of the stem cell research industry in perspective of natural bioreactor and producer cells. Table 6 summarizes some of these growth factors and cytokines which produced by several companies that are used for laboratory and research purposes. It is noteworthy that the GMP of MSCs-derived growth factors and cytokines are different from those stem cells that transplanted to patients. When MSC-derived growth factors and cytokines are packaged properly, they can be transported easily like other drugs and they do not need cryopreservation. However, in comparison with recombinant growth factors that may be stable for a long period of time and also produced on a larger scale in non-stem cell hosts, MSC-derived growth factors and cytokines which need more optimization in terms of production and stability. A number of growth factors and cytokines in the secretome which have been expressed separately in MSCs using recombinant DNA technology are presented in Table 7. For clinical application, a large amount of growth factors/cytokines are needed. Hence, manufacturing large quantities (scale-up) of hMSC's secretome based on GMPprocedures will be challenging.

For clinical application, it is essential to improve the production of MSC-derived growth factors/ cytokines. A variety of biotechnological techniques such as suspension culture, cultivating with three-dimensional (3D) scaffolds, cultivating with an advanced bioreactor, cultivating under sublethal doses of oxidative stress, hypoxia and magnetic field (MF) can be employed in this area [16, 143]. The aims of aforementioned techniques are to simulate and reproduce the stem cell niche in order to improve MSCs quality and in turn MSC-derived growth factors/cytokines. In addition, genetically modified MSCs with cytoprotective factors such as nuclear factor-

 Table 7
 A number of growth factors and cytokines in the secretome which have separately been expressed in MSCs by recombinant DNA technology

GMP Grade MSC Agents	
GMP Recombinant Human FGF-2	GMP Recombinant Human IL-2
GMP Recombinant Human Flt3-ligand	GMP Recombinant Human IL-3
GMP Recombinant Human GM-CSF	GMP Recombinant Human IL-4
GMP Recombinant Human GM-CSF	GMP Recombinant Human IL-6
GMP Recombinant Human SCF	GMP Recombinant Human IL-7
GMP Recombinant Human TNF- $\alpha$	GMP Recombinant Human IL-15
GMP Recombinant Human IL-1 $\beta$	GMP Recombinant Human IL-21

erythroid 2-related factor 2 (Nrf2), TGF $\beta$ 1, HO-1, lipocalin 2 (Lcn2), VEGF, hypoxia-inducible factor (HIF-1 $\alpha$ ), IGF-1 and etc. could be other strategies to improve MSC-derived growth factors/ cytokines [16–18, 76].

## Conclusion

In clinical perspective, MSCs have drawn lots of interests for more than one decade. However, concerns about tumor formation and low survival rate after transplantation are the main limitations that impair their widespread usage in clinic. Instead, a number of studies have shown that MSCs can exert their therapeutic roles via producing of a vast array of bioactive molecules such as growth factors, cytokines, peptides, hormones, and microRNAs. These unique properties of MSCs are convincing to call MSCs as an arsenal of therapeutic agents. In other words, MSCs naturally and innately act as a bioreactor in order to produce a large number of valuable pharmaceutical products as well as, open a new horizon for basic, clinical scientists and marketing.

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### **Compliance with Ethical Standards**

Conflict of Interest The authors declare there is no conflict of interest.

#### References

- Youssef A, Aboalola D, & Han VK. (2017). The roles of insulinlike growth factors in mesenchymal stem cell niche. Stem Cells International;2017.
- Dominici, M., Le Blanc, K., Mueller, I., et al. (2006). Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy*, 8, 315–317.
- Vizoso, F. J., Eiro, N., Cid, S., Schneider, J., & Perez-Fernandez, R. (2017). Mesenchymal stem cell secretome: Toward cell-free

therapeutic strategies in regenerative medicine. *International Journal of Molecular Sciences*, 18, 1852.

- Kim, H. O., Choi, S.-M., & Kim, H.-S. (2013). Mesenchymal stem cell-derived secretome and microvesicles as a cell-free therapeutics for neurodegenerative disorders. *Tissue Engineering and Regenerative Medicine*, 10, 93–101.
- Ding, D.-C., Shyu, W.-C., & Lin, S.-Z. (2011). Mesenchymal stem cells. *Cell Transplantation*, 20, 5–14.
- Wakao, S., Kuroda, Y., Ogura, F., Shigemoto, T., & Dezawa, M. (2012). Regenerative effects of mesenchymal stem cells: contribution of muse cells, a novel pluripotent stem cell type that resides in mesenchymal cells. *Cell*, *1*, 1045–1060.
- Koh, M. B., & Suck, G. (2012). Cell therapy: promise fulfilled? Biologicals, 40, 214–217.
- Phinney, D. G., & Pittenger, M. F. (2017). Concise review: MSCderived exosomes for cell-free therapy. *Stem Cells*, 35, 851–858.
- Schoeberlein, A., Mueller, M., Reinhart, U., Sager, R., Messerli, M., & Surbek, D. V. (2011). Homing of placenta-derived mesenchymal stem cells after perinatal intracerebral transplantation in a rat model. *American Journal of Obstetrics & Gynecology*, 205(277), e1–e6.
- Mueller, M., Wolfs, T. G., Schoeberlein, A., Gavilanes, A. W., Surbek, D., & Kramer, B. W. (2016). Mesenchymal stem/ stromal cells—a key mediator for regeneration after perinatal morbidity? *Molecular and Cellular Pediatrics*, *3*, 6.
- Lee, K. A., Shim, W., Paik, M. J., et al. (2009). Analysis of changes in the viability and gene expression profiles of human mesenchymal stromal cells over time. *Cytotherapy*, 11, 688–697.
- Han, S.-M., Han, S.-H., Coh, Y.-R., et al. (2014). Enhanced proliferation and differentiation of Oct4-and Sox2-overexpressing human adipose tissue mesenchymal stem cells. *Experimental & Molecular Medicine*, 46, e101.
- Hagberg, H., Mallard, C., Ferriero, D. M., et al. (2015). The role of inflammation in perinatal brain injury. *Nature Reviews Neurology*, 11, 192.
- Francois, S., Mouiseddine, M., Allenet-Lepage, B., et al. (2013). Human mesenchymal stem cells provide protection against radiation-induced liver injury by antioxidative process, vasculature protection, hepatocyte differentiation, and trophic effects. *BioMed Research International, 2013.*
- 15. Fossett, E., & Khan, W. (2012). Optimising human mesenchymal stem cell numbers for clinical application: a literature review. *Stem Cells International, 2012.*
- Amiri, F., Jahanian-Najafabadi, A., & Roudkenar, M. H. (2015). In vitro augmentation of mesenchymal stem cells viability in stressful microenvironments. *Cell Stress and Chaperones*, 20, 237–251.
- Halabian, R., Tehrani, H. A., Jahanian-Najafabadi, A., & Roudkenar, M. H. (2013). Lipocalin-2-mediated upregulation of various antioxidants and growth factors protects bone marrowderived mesenchymal stem cells against unfavorable microenvironments. *Cell stress and chaperones*, 18, 785–800.
- Kiani, A. A., Kazemi, A., Halabian, R., Mohammadipour, M., Jahanian-Najafabadi, A., & Roudkenar, M. H. (2013). HIF-1α confers resistance to induced stress in bone marrow-derived mesenchymal stem cells. *Archives of Medical Research*, 44, 185–193.
- Sotiropoulou, P. A., Perez, S. A., Salagianni, M., Baxevanis, C. N., & Papamichail, M. (2006). Characterization of the optimal culture conditions for clinical scale production of human mesenchymal stem cells. *Stem Cells*, *24*, 462–471.
- Duggal, S., & Brinchmann, J. E. (2011). Importance of serum source for the in vitro replicative senescence of human bone marrow derived mesenchymal stem cells. *Journal of Cellular Physiology*, 226, 2908–2915.
- Kornicka K, Marycz K, Tomaszewski KA, Marędziak M, Śmieszek A. (2015). The effect of age on osteogenic and

adipogenic differentiation potential of human adipose derived stromal stem cells (hASCs) and the impact of stress factors in the course of the differentiation process. Oxidative Medicine and Cellular Longevity ;2015.

- Bertolo, A., Capossela, S., Fränkl, G., Baur, M., Pötzel, T., & Stoyanov, J. (2017). Oxidative status predicts quality in human mesenchymal stem cells. *Stem Cell Research & Therapy*, 8, 3.
- Maredziak, M., Marycz, K., Tomaszewski, K. A., Kornicka, K., & Henry, B. M. (2016). The influence of aging on the regenerative potential of human adipose derived mesenchymal stem cells. *Stem Cells International*, 2016, 2152435.
- Sell, S. (2010). On the stem cell origin of cancer. *The American Journal of Pathology*, 176, 2584–2594.
- Kucia, M., Reca, R., Miekus, K., et al. (2005). Trafficking of normal stem cells and metastasis of cancer stem cells involve similar mechanisms: pivotal role of the SDF-1–CXCR4 axis. *Stem Cells, 23*, 879–894.
- Marycz K, Kornicka K, Grzesiak J, Śmieszek A, Szłapka J. (2016). Macroautophagy and selective mitophagy ameliorate chondrogenic differentiation potential in adipose stem cells of equine metabolic syndrome: New findings in the field of progenitor cells differentiation. Oxidative Medicine and Cellular Longevity ;2016.
- Yang, D., Wang, W., Li, L., et al. (2013). The relative contribution of paracine effect versus direct differentiation on adipose-derived stem cell transplantation mediated cardiac repair. *PLoS One*, *8*, e59020.
- Prockop, D. (2007). "Stemness" does not explain the repair of many tissues by mesenchymal stem/multipotent stromal cells (MSCs). *Clinical Pharmacology & Therapeutics*, 82, 241–243.
- Bai, L., Shao, H., Wang, H., et al. (2017). Effects of mesenchymal stem cell-derived exosomes on experimental autoimmune uveitis. *Scientific Reports*, 7, 4323.
- Bruno, S., Grange, C., Deregibus, M. C., et al. (2009). Mesenchymal stem cell-derived microvesicles protect against acute tubular injury. *Journal of the American Society of Nephrology*, 20, 1053–1067.
- Bruno, S., Grange, C., Collino, F., et al. (2012). Microvesicles derived from mesenchymal stem cells enhance survival in a lethal model of acute kidney injury. *PLoS One*, 7, e33115.
- Aliotta, J. M., Pereira, M., Wen, S., et al. (2016). Exosomes induce and reverse monocrotaline-induced pulmonary hypertension in mice. *Cardiovascular Research*, *110*, 319–330.
- Lee C, Mitsialis SA, Aslam M, et al. (2012). Exosomes mediate the cytoprotective action of mesenchymal stromal cells on hypoxia-induced pulmonary hypertension. Circulation: Circulationaha. 112.114173.
- 34. Zhang, B., Wang, M., Gong, A., et al. (2015). HucMSC-exosome mediated-Wnt4 signaling is required for cutaneous wound healing. *Stem Cells*, *33*, 2158–2168.
- Zhang, B., Wu, X., Zhang, X., et al. (2015). Human umbilical cord mesenchymal stem cell exosomes enhance angiogenesis through the Wnt4/β-catenin pathway. *Stem Cells Translational Medicine*, 4, 513–522.
- Bermudez, M. A., Sendon-Lago, J., Eiro, N., et al. (2015). Corneal epithelial wound healing and bactericidal effect of conditioned medium from human uterine cervical stem cells. *Investigative Ophthalmology & Visual Science*, 56, 983–992.
- Bhang, S. H., Lee, S., Shin, J.-Y., Lee, T.-J., Jang, H.-K., & Kim, B.-S. (2014). Efficacious and clinically relevant conditioned medium of human adipose-derived stem cells for therapeutic angiogenesis. *Molecular Therapy*, 22, 862–872.
- Zagoura DS, Roubelakis MG, Bitsika V, et al. (2011). Therapeutic potential of a distinct population of human amniotic fluid mesenchymal stem cells and their secreted molecules in mice with acute hepatic failure. Gut:gutjnl-2011-300908.

- Park, B.-S., Kim, W.-S., Choi, J.-S., et al. (2010). Hair growth stimulated by conditioned medium of adipose-derived stem cells is enhanced by hypoxia: evidence of increased growth factor secretion. *Biomedical Research*, 31, 27–34.
- Mirabella, T., Cilli, M., Carlone, S., Cancedda, R., & Gentili, C. (2011). Amniotic liquid derived stem cells as reservoir of secreted angiogenic factors capable of stimulating neo-arteriogenesis in an ischemic model. *Biomaterials*, 32, 3689–3699.
- Lee, M. J., Kim, J., Lee, K. I., Shin, J. M., Chae, J. I., & Chung, H. M. (2011). Enhancement of wound healing by secretory factors of endothelial precursor cells derived from human embryonic stem cells. *Cytotherapy*, *13*, 165–178.
- Kim, J., Lee, J. H., Yeo, S. M., Chung, H. M., & Chae, J.-I. (2014). Stem cell recruitment factors secreted from cord blood-derived stem cells that are not secreted from mature endothelial cells enhance wound healing. *In Vitro Cellular & Developmental Biology-Animal*, 50, 146–154.
- 43. Ray, P., Devaux, Y., Stolz, D. B., et al. (2003). Inducible expression of keratinocyte growth factor (KGF) in mice inhibits lung epithelial cell death induced by hyperoxia. *Proceedings of the National Academy of Sciences, 100*, 6098–6103.
- Turner J-E, Morrison PJ, Wilhelm C, et al. (2013). IL-9–mediated survival of type 2 innate lymphoid cells promotes damage control in helminth-induced lung inflammation. *Journal of Experimental Medicine* :jem. 20130071.
- Bakondi, B., Shimada, I. S., Perry, A., et al. (2009). CD133 identifies a human bone marrow stem/progenitor cell sub-population with a repertoire of secreted factors that protect against stroke. *Molecular Therapy*, 17, 1938–1947.
- Martinez-Agosto, J. A., Mikkola, H. K., Hartenstein, V., & Banerjee, U. (2007). The hematopoietic stem cell and its niche: a comparative view. *Genes & Development*, *21*, 3044–3060.
- Beer L, Mildner M, & Ankersmit HJ. (2017). Cell secretome based drug substances in regenerative medicine: when regulatory affairs meet basic science. *Annals of Translational Medicine* ;5.
- 48. Pawitan JA. (2014). Prospect of stem cell conditioned medium in regenerative medicine. *BioMed Research International* ;2014.
- Cantinieaux, D., Quertainmont, R., Blacher, S., et al. (2013). Conditioned medium from bone marrow-derived mesenchymal stem cells improves recovery after spinal cord injury in rats: an original strategy to avoid cell transplantation. *PLoS One, 8*, e69515.
- Bermudez, M. A., Sendon-Lago, J., Seoane, S., et al. (2016). Antiinflammatory effect of conditioned medium from human uterine cervical stem cells in uveitis. *Experimental Eye Research*, 149, 84–92.
- Chang, C.-P., Chio, C.-C., Cheong, C.-U., Chao, C.-M., Cheng, B.-C., & Lin, M.-T. (2013). Hypoxic preconditioning enhances the therapeutic potential of the secretome from cultured human mesenchymal stem cells in experimental traumatic brain injury. *Clinical Science*, 124, 165–176.
- Ionescu, L., Byrne, R. N., van Haaften, T., et al. (2012). Stem cell conditioned medium improves acute lung injury in mice: in vivo evidence for stem cell paracrine action. *American Journal of Physiology-Lung Cellular and Molecular Physiology, 303*, L967–LL77.
- Katagiri, W., Watanabe, J., Toyama, N., Osugi, M., Sakaguchi, K., & Hibi, H. (2017). Clinical study of bone regeneration by conditioned medium from mesenchymal stem cells after maxillary sinus floor elevation. *Implant Dentistry*, 26, 607–612.
- Legaki, E., Roubelakis, M., Theodoropoulos, G., et al. (2016). Therapeutic potential of secreted molecules derived from human amniotic fluid mesenchymal stem/stroma cells in a mice model of colitis. *Stem Cell Reviews and Reports, 12*, 604–612.
- 55. Sevivas, N., Teixeira, F. G., Portugal, R., et al. (2017). Mesenchymal stem cell secretome: a potential tool for the prevention of muscle degenerative changes associated with chronic

rotator cuff tears. *The American Journal of Sports Medicine, 45*, 179–188.

- Timmers, L., Lim, S. K., Arslan, F., et al. (2008). Reduction of myocardial infarct size by human mesenchymal stem cell conditioned medium. *Stem Cell Research*, *1*, 129–137.
- Zhou B-R, Xu Y, Guo S-L, et al. (2013). The effect of conditioned media of adipose-derived stem cells on wound healing after ablative fractional carbon dioxide laser resurfacing. *BioMed Research International* ;2013.
- Eiró, N., Sendon-Lago, J., Seoane, S., et al. (2014). Potential therapeutic effect of the secretome from human uterine cervical stem cells against both cancer and stromal cells compared with adipose tissue stem cells. *Oncotarget*, 5, 10692.
- Ozbey, G., Gorczynski, R., & Erin, N. (2014). Stability of cytokines in supernatants of stimulated mouse immune cells. *European Cytokine Network*, 25, 30–34.
- Lai, R. C., Arslan, F., Lee, M. M., et al. (2010). Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. *Stem Cell Research*, *4*, 214–222.
- Yu, B., Kim, H. W., Gong, M., et al. (2015). Exosomes secreted from GATA-4 overexpressing mesenchymal stem cells serve as a reservoir of anti-apoptotic microRNAs for cardioprotection. *International Journal of Cardiology*, 182, 349–360.
- Xin, H., Li, Y., Cui, Y., Yang, J. J., Zhang, Z. G., & Chopp, M. (2013). Systemic administration of exosomes released from mesenchymal stromal cells promote functional recovery and neurovascular plasticity after stroke in rats. *Journal of Cerebral Blood Flow & Metabolism, 33*, 1711–1715.
- 63. Ophelders, D. R., Wolfs, T. G., Jellema, R. K., et al. (2016). Mesenchymal Stromal Cell-Derived Extracellular Vesicles Protect the Fetal Brain After Hypoxia-Ischemia. *Stem Cells Translational Medicine*, *5*, 754–763.
- Hu, G.-w., Li, Q., Niu, X., et al. (2015). Exosomes secreted by human-induced pluripotent stem cell-derived mesenchymal stem cells attenuate limb ischemia by promoting angiogenesis in mice. *Stem Cell Research & Therapy*, *6*, 10.
- Li, T., Yan, Y., Wang, B., et al. (2012). Exosomes derived from human umbilical cord mesenchymal stem cells alleviate liver fibrosis. *Stem Cells and Development*, *22*, 845–854.
- Tan, C. Y., Lai, R. C., Wong, W., Dan, Y. Y., Lim, S.-K., & Ho, H. K. (2014). Mesenchymal stem cell-derived exosomes promote hepatic regeneration in drug-induced liver injury models. *Stem Cell Research & Therapy*, 5, 76.
- Rager, T. M., Olson, J. K., Zhou, Y., Wang, Y., & Besner, G. E. (2016). Exosomes secreted from bone marrow-derived mesenchymal stem cells protect the intestines from experimental necrotizing enterocolitis. *Journal of Pediatric Surgery*, *51*, 942–947.
- 68. Monsel, A., Zhu, Y.-g., Gennai, S., et al. (2015). Therapeutic effects of human mesenchymal stem cell–derived microvesicles in severe pneumonia in mice. *American Journal of Respiratory and Critical Care Medicine, 192*, 324–336.
- Gennai, S., Monsel, A., Hao, Q., Park, J., Matthay, M., & Lee, J. (2015). Microvesicles derived from human mesenchymal stem cells restore alveolar fluid clearance in human lungs rejected for transplantation. *American Journal of Transplantation*, 15, 2404–2412.
- Choi, M., Ban, T., & Rhim, T. (2014). Therapeutic use of stem cell transplantation for cell replacement or cytoprotective effect of microvesicle released from mesenchymal stem cell. *Molecules* and Cells, 37, 133.
- Zhang, J., Guan, J., Niu, X., et al. (2015). Exosomes released from human induced pluripotent stem cells-derived MSCs facilitate cutaneous wound healing by promoting collagen synthesis and angiogenesis. *Journal of Translational Medicine*, 13, 49.
- 72. Fang, S., Xu, C., Zhang, Y., et al. (2016). Umbilical cord-derived mesenchymal stem cell-derived exosomal microRNAs suppress myofibroblast differentiation by inhibiting the transforming

growth factor-β/SMAD2 pathway during wound healing. *Stem Cells Translational Medicine*, *5*, 1425–1439.

- Nakamura, Y., Miyaki, S., Ishitobi, H., et al. (2015). Mesenchymal-stem-cell-derived exosomes accelerate skeletal muscle regeneration. *FEBS Letters*, 589, 1257–1265.
- Barry, F., & Murphy, M. (2013). Mesenchymal stem cells in joint disease and repair. *Nature Reviews Rheumatology*, 9, 584.
- Aghajani Nargesi, A., O Lerman, L., & Eirin, A. (2017). Mesenchymal stem cell-derived extracellular vesicles for renal repair. *Current Gene Therapy*, 17, 29–42.
- Roushandeh, A. M., Bahadori, M., & Roudkenar, M. H. (2017). Mesenchymal Stem Cell-based Therapy as a New Horizon for Kidney Injuries. *Archives of Medical Research*, 48, 133–146.
- 77. Halabian, R., Roudkenar, M. H., Jahanian-Najafabadi, A., Hosseini, K. M., & Tehrani, H. A. (2015). Co-culture of bone marrow-derived mesenchymal stem cells overexpressing lipocalin 2 with HK-2 and HEK293 cells protects the kidney cells against cisplatin-induced injury. *Cell Biology International*, 39, 152–163.
- Kraitchman, D. L., Tatsumi, M., Gilson, W. D., et al. (2005). Dynamic imaging of allogeneic mesenchymal stem cells trafficking to myocardial infarction. *Circulation*, *112*, 1451–1461.
- Bittira, B., Shum-Tim, D., Al-Khaldi, A., & Chiu, R. C. (2003). Mobilization and homing of bone marrow stromal cells in myocardial infarction. *European Journal of Cardio-Thoracic Surgery*, 24, 393–398.
- Shake, J. G., Gruber, P. J., Baumgartner, W. A., et al. (2002). Mesenchymal stem cell implantation in a swine myocardial infarct model: engraftment and functional effects. *The Annals of Thoracic Surgery*, 73, 1919–1926.
- Fontaine, M. J., Shih, H., Schäfer, R., & Pittenger, M. F. (2016). Unraveling the mesenchymal stromal cells' paracrine immunomodulatory effects. *Transfusion Medicine Reviews*, 30, 37–43.
- Aslam, M., Baveja, R., Liang, O. D., et al. (2009). Bone marrow stromal cells attenuate lung injury in a murine model of neonatal chronic lung disease. *American Journal of Respiratory and Critical Care Medicine*, 180, 1122–1130.
- Ho, J. C., Lai, W. H., Li, M. F., et al. (2012). Reversal of endothelial progenitor cell dysfunction in patients with type 2 diabetes using a conditioned medium of human embryonic stem cellderived endothelial cells. *Diabetes/Metabolism Research and Reviews, 28*, 462–473.
- Di Santo, S., Yang, Z., von Ballmoos, M. W., et al. (2009). Novel cell-free strategy for therapeutic angiogenesis: in vitro generated conditioned medium can replace progenitor cell transplantation. *PLoS One, 4*, e5643.
- Inukai, T., Katagiri, W., Yoshimi, R., et al. (2013). Novel application of stem cell-derived factors for periodontal regeneration. *Biochemical and Biophysical Research Communications*, 430, 763–768.
- Sadat, S., Gehmert, S., Song, Y.-H., et al. (2007). The cardioprotective effect of mesenchymal stem cells is mediated by IGF-I and VEGF. *Biochemical and Biophysical Research Communications*, 363, 674–679.
- See, F., Seki, T., Psaltis, P. J., et al. (2011). Therapeutic effects of human STRO-3-selected mesenchymal precursor cells and their soluble factors in experimental myocardial ischemia. *Journal of Cellular and Molecular Medicine*, 15, 2117–2129.
- Cho, Y. J., Song, H. S., Bhang, S., et al. (2012). Therapeutic effects of human adipose stem cell-conditioned medium on stroke. *Journal of Neuroscience Research*, 90, 1794–1802.
- Sze, S. K., de Kleijn, D. P., Lai, R. C., et al. (2007). Elucidating the secretion proteome of human embryonic stem cell-derived mesenchymal stem cells. *Molecular & Cellular Proteomics*, 6, 1680–1689.
- Xiong, L.-L., Liu, F., Lu, B.-T., et al. (2017). Bone Marrow Mesenchymal Stem-Cell Transplantation Promotes Functional Improvement Associated with CNTF-STAT3 Activation after

Hemi-Sectioned Spinal Cord Injury in Tree Shrews. *Frontiers in Cellular Neuroscience*, 11, 172.

- Chen, L., Tredget, E. E., Wu, P. Y., & Wu, Y. (2008). Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. *PLoS One*, *3*, e1886.
- 92. Gao, F., Chiu, S., Motan, D., et al. (2017). Mesenchymal stem cells and immunomodulation: current status and future prospects. *Cell Death & Disease*, *7*, e2062.
- Ruvolo P, Qiu Y, Ruvolo V, et al. (2015). Role of Mesenchymal Stem Cell Galectin 3 in the AML Tumor Microenvironment. Am Soc Hematology.
- 94. Ling, W., Zhang, J., & Yuan, Z., et al. (2014). Mesenchymal stem cells use IDO to regulate immunity in tumor microenvironment. *Cancer Research*.
- Lourenco, S., Teixeira, V. H., Kalber, T., Jose, R. J., Floto, R. A., & Janes, S. M. (2015). Macrophage migration inhibitory factor– CXCR4 is the dominant chemotactic axis in human mesenchymal stem cell recruitment to tumors. *The Journal of Immunology, 194*, 3463–3474.
- Kim, H. S., Yun, J. W., Shin, T. H., et al. (2015). Human umbilical cord blood mesenchymal stem cell-derived PGE2 and TGF-β1 alleviate atopic dermatitis by reducing mast cell degranulation. *Stem Cells*, 33, 1254–1266.
- Lee, R. H., Pulin, A. A., Seo, M. J., et al. (2009). Intravenous hMSCs improve myocardial infarction in mice because cells embolized in lung are activated to secrete the anti-inflammatory protein TSG-6. *Cell Stem Cell*, 5, 54–63.
- Qi, Y., Jiang, D., Sindrilaru, A., et al. (2014). TSG-6 released from intradermally injected mesenchymal stem cells accelerates wound healing and reduces tissue fibrosis in murine full-thickness skin wounds. *Journal of Investigative Dermatology*, 134, 526–537.
- Krasnodembskaya, A., Song, Y., Fang, X., et al. (2010). Antibacterial effect of human mesenchymal stem cells is mediated in part from secretion of the antimicrobial peptide LL-37. *Stem Cells*, 28, 2229–2238.
- Németh, K., Leelahavanichkul, A., Yuen, P. S., et al. (2009). Bone marrow stromal cells attenuate sepsis via prostaglandin E 2–dependent reprogramming of host macrophages to increase their interleukin-10 production. *Nature Medicine*, 15, 42.
- Choi, H., Lee, R. H., Bazhanov, N., Oh, J. Y., & Prockop, D. J. (2011). Anti-inflammatory protein TSG-6 secreted by activated MSCs attenuates zymosan-induced mouse peritonitis by decreasing TLR2/NF-κB signaling in resident macrophages. *Blood*, *118*, 330–338.
- Keating, A. (2012). Mesenchymal stromal cells: new directions. Cell Stem Cell, 10, 709–716.
- Fierabracci, A., Del Fattore, A., Muraca, M., Vittorio Delfino, D., & Muraca, M. (2016). The use of mesenchymal stem cells for the treatment of autoimmunity: from animals models to human disease. *Current Drug Targets*, 17, 229–238.
- Nauta, A. J., Kruisselbrink, A. B., Lurvink, E., Willemze, R., & Fibbe, W. E. (2006). Mesenchymal stem cells inhibit generation and function of both CD34+–derived and monocyte-derived dendritic cells. *The Journal of Immunology*, *177*, 2080–2087.
- English, K., Barry, F. P., & Mahon, B. P. (2008). Murine mesenchymal stem cells suppress dendritic cell migration, maturation and antigen presentation. *Immunology Letters*, 115, 50–58.
- Spaggiari, G. M., Abdelrazik, H., Becchetti, F., & Moretta, L. (2009). MSCs inhibit monocyte-derived DC maturation and function by selectively interfering with the generation of immature DCs: central role of MSC-derived prostaglandin E2. *Blood*, *113*, 6576–6583.
- 107. Li, Y.-P., Paczesny, S., Lauret, E., et al. (2008). Human mesenchymal stem cells license adult CD34+ hemopoietic progenitor cells to differentiate into regulatory dendritic cells through activation of the Notch pathway. *The Journal of Immunology, 180*, 1598–1608.

- 108. Zhang, B., Liu, R., Shi, D., et al. (2009). Mesenchymal stem cells induce mature dendritic cells into a novel Jagged-2–dependent regulatory dendritic cell population. *Blood*, *113*, 46–57.
- Li, Q., Fang, Y., Li, X., et al. (2013). Mechanism of the plant cytochrome P450 for herbicide resistance: a modelling study. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 28, 1182–1191.
- Liu, X., Qu, X., Chen, Y., et al. (2012). Mesenchymal stem/ stromal cells induce the generation of novel IL-10–dependent regulatory dendritic cells by SOCS3 activation. *The Journal of Immunology, 189*, 1182–1192.
- Schu, S., Nosov, M., O'Flynn, L., et al. (2012). Immunogenicity of allogeneic mesenchymal stem cells. *Journal of Cellular and Molecular Medicine*, 16, 2094–2103.
- 112. Le Blanc, K., Rasmusson, I., Sundberg, B., et al. (2004). Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *The Lancet, 363*, 1439–1441.
- 113. English, K., French, A., & Wood, K. J. (2010). Mesenchymal stromal cells: facilitators of successful transplantation? *Cell Stem Cell*, *7*, 431–442.
- Prigozhina, T. B., Khitrin, S., Elkin, G., Eizik, O., Morecki, S., & Slavin, S. (2008). Mesenchymal stromal cells lose their immunosuppressive potential after allotransplantation. *Experimental Hematology*, 36, 1370–1376.
- 115. Sudres, M., Norol, F., Trenado, A., et al. (2006). Bone marrow mesenchymal stem cells suppress lymphocyte proliferation in vitro but fail to prevent graft-versus-host disease in mice. *The Journal of Immunology*, *176*, 7761–7767.
- Baylink, D. J., Finkelman, R. D., & Mohan, S. (1993). Growth factors to stimulate bone formation. *Journal of Bone and Mineral Research*, 8.
- 117. Shim, K. S. (2015). Pubertal growth and epiphyseal fusion. *Annals of Pediatric Endocrinology & metabolism, 20*, 8–12.
- 118. Urist, M. R. (1965). Bone: formation by autoinduction. *Science*, *150*, 893–899.
- 119. Astori, G., Vignati, F., Bardelli, S., et al. (2007). " In vitro" and multicolor phenotypic characterization of cell subpopulations identified in fresh human adipose tissue stromal vascular fraction and in the derived mesenchymal stem cells. *Journal of Translational Medicine*, *5*, 55.
- Guo, J., Nguyen, A., Banyard, D. A., et al. (2016). Stromal vascular fraction: a regenerative reality? Part 2: Mechanisms of regenerative action. *Journal of Plastic, Reconstructive & Aesthetic Surgery, 69,* 180–188.
- Schouten, H. (2006). Neutropenia management. Annals of Oncology, 17, x85-xx9.
- 122. James, R., & Kinsey, S. (2006). The investigation and management of chronic neutropenia in children. *Archives of Disease in Childhood, 91*, 852–858.
- 123. Jubelirer, S. J. (2011). The benefit of the neutropenic diet: fact or fiction? *The Oncologist, 16*, 704–707.
- Steed, D. L. (1997). The role of growth factors in wound healing. Surgical Clinics of North America, 77, 575–586.
- 125. Bermudez, M. A., Sendon-Lago, J., Eiro, N., et al. (2015). Corneal epithelial wound healing and bactericidal effect of conditioned medium from human uterine cervical stem cells effect of CMhUCESCs on wound healing in dry eye. *Investigative Ophthalmology & Visual Science*, 56, 983–992.
- Ranganath, S. H., Levy, O., Inamdar, M. S., & Karp, J. M. (2012). Harnessing the mesenchymal stem cell secretome for the treatment of cardiovascular disease. *Cell Stem Cell*, 10, 244–258.

- 127. Pankajakshan D, & Agrawal DK. (2014). Mesenchymal stem cell paracrine factors in vascular repair and regeneration. *Journal of Biomedical Technology and Research* 1.
- Du, Z., Wei, C., Cheng, K., et al. (2013). Mesenchymal stem cellconditioned medium reduces liver injury and enhances regeneration in reduced-size rat liver transplantation. *Journal of Surgical Research*, 183, 907–915.
- Forte, A., Finicelli, M., Mattia, M., et al. (2008). Mesenchymal stem cells effectively reduce surgically induced stenosis in rat carotids. *Journal of Cellular Physiology*, 217, 789–799.
- Forte, A., Rinaldi, B., Sodano, L., et al. (2012). Stem cell therapy for arterial restenosis: potential parameters contributing to the success of bone marrow-derived mesenchymal stromal cells. *Cardiovascular Drugs and Therapy*, 26, 9–21.
- 131. Shoji, M., Oskowitz, A., Malone, C. D., Prockop, D. J., & Pochampally, R. (2011). Human mesenchymal stromal cells (MSCs) reduce neointimal hyperplasia in a mouse model of flow-restriction by transient suppression of anti-inflammatory cytokines. *Journal of Atherosclerosis and Thrombosis, 18*, 464–474.
- 132. Takahashi, M., Suzuki, E., Oba, S., et al. (2009). Adipose tissuederived stem cells inhibit neointimal formation in a paracrine fashion in rat femoral artery. *American Journal of Physiology-Heart and Circulatory Physiology, 298*, H415–HH23.
- Sato, K., Ozaki, K., Oh, I., et al. (2007). Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells. *Blood*, 109, 228–234.
- Wang, C.-H., Cherng, W.-J., Yang, N.-I., et al. (2008). Lateoutgrowth endothelial cells attenuate intimal hyperplasia contributed by mesenchymal stem cells after vascular injury. *Arteriosclerosis, Thrombosis, and Vascular Biology, 28*, 54–60.
- Shoji, M., Koba, S., & Kobayashi, Y. (2014). Roles of bone-marrowderived cells and inflammatory cytokines in neointimal hyperplasia after vascular injury. *BioMed Research International* 2014.
- 136. Marędziak, M., Marycz, K., Lewandowski, D., Siudzińska, A., & Śmieszek, A. (2015). Static magnetic field enhances synthesis and secretion of membrane-derived microvesicles (MVs) rich in VEGF and BMP-2 in equine adipose-derived stromal cells (EqASCs)—a new approach in veterinary regenerative medicine. *In Vitro Cellular & Developmental Biology-Animal*, 51, 230–240.
- 137. Rajan, T. S., Giacoppo, S., Diomede, F., et al. (2016). The secretome of periodontal ligament stem cells from MS patients protects against EAE. *Scientific Reports*, 6, 38743.
- Teixeira, F. G., Carvalho, M. M., Panchalingam, K. M., et al. (2017). Impact of the secretome of human mesenchymal stem cells on brain structure and animal behavior in a rat model of Parkinson's disease. *Stem Cells Translational Medicine*, *6*, 634–646.
- Kim, M. J., Kim, Z.-H., Kim, S.-M., & Choi, Y.-S. (2016). Conditioned medium derived from umbilical cord mesenchymal stem cells regenerates atrophied muscles. *Tissue and Cell*, 48, 533–543.
- 140. Qi, K., Li, N., Zhang, Z., & Melino, G. (2017). Tissue regeneration: The crosstalk between mesenchymal stem cells and immune response. *Cellular Immunology*.
- Duplantier, A. J., & van Hoek, M. L. (2013). The human cathelicidin antimicrobial peptide LL-37 as a potential treatment for polymicrobial infected wounds. *Frontiers in Immunology*, 4, 143.
- Yue, R., Zhou, B. O., Shimada, I. S., Zhao, Z., & Morrison, S. J. (2016). Leptin receptor promotes adipogenesis and reduces osteogenesis by regulating mesenchymal stromal cells in adult bone marrow. *Cell Stem Cell*, *18*, 782–796.
- 143. Li, J., Peng, X., Zeng, X., et al. (2015). Estrogen secreted by mesenchymal stem cells necessarily determines their feasibility of therapeutical application. *Scientific Reports*, 5, 15286.